

WHAT IS CLAIMED IS:

- 1           1. A nucleic acid extraction device, comprising:  
2           a body having at least one chamber with at least one  
3 inlet channel; and  
4           a porous flow-through plug disposed within the chamber,  
5 the plug having nucleic acid binding properties.
- 1           2. The nucleic acid extraction device of claim 1,  
2 wherein said chamber has a width in the range of 0.05 to  
3 2.0mm.
- 1           3. The nucleic acid extraction device of claim 2,  
2 wherein said chamber has a width in the range of 0.1 to 0.5mm.
- 1           4. The nucleic acid extraction device of claim 3,  
2 wherein said chamber has a depth in the range of 0.05 to 1mm.
- 1           5. The nucleic acid extraction device of claim 1,  
2 wherein said plug is a deformable plug.
- 1           6. The nucleic acid extraction device of claim 1,  
2 wherein the plug comprises glass wool.
- 1           7. The nucleic acid extraction device of claim 5,  
2 wherein the plug comprises glass wool.
- 1           8. A nucleic acid extraction device, comprising:  
2           a body having at least one chamber and at least one  
3 inlet channel; and  
4           a textured surface disposed within the chamber, the  
5 surface having nucleic acid binding properties.
- 1           9. A nucleic acid extraction device, comprising:  
2           a body having at least one chamber and at least one  
3 inlet channel; and

4 an affinity surface having particles attached thereto,  
5 the particles having nucleic acid binding properties.

1 10. The device of claim 1, wherein the plug is  
2 pretreated with an agent for enhancing the nucleic acid  
3 binding properties.

1 11. The device of claim 10, wherein said agent is  
2 selected from the group consisting of acids, bases, silanes,  
3 polysine, tethered antibodies, synthesized nucleic acids, and  
4 Poly-T DNA.

1 12. The device of claim 10, wherein the structure is  
2 an open cell foam.

3 13. The nucleic acid extraction device of claim 5,  
4 further comprising:  
5 a flexible diaphragm for compressing said plug thereby  
6 removing trapped liquids.

1 14. The nucleic acid extraction device of claim 13,  
2 wherein  
3 the flexible diaphragm is disposed between a pneumatic  
4 port and the structure, the device further comprising a  
5 pressure system for displacing the flexible diaphragm to draw  
6 a sample through the inlet channel into the chamber.

1 15. The nucleic acid extraction device of claim 1,  
2 wherein said structure is an affinity surface in a flow  
3 through chamber.

1 16. The nucleic acid extraction device of claim 9,  
2 wherein said affinity surface has controlled-pore glass  
3 structures attached thereto.

1 17. The nucleic acid extraction device of claim 9,  
2 wherein said affinity surface has glass spheres attached  
3 thereto.

1 18. The nucleic acid extraction device of claim 9,  
2 wherein said affinity surface has cellulose particles attached  
3 thereto.

1 19. The nucleic acid extraction device of claim 8,  
2 wherein said affinity surface is microfabricated.

1 20. The nucleic acid extraction device of claim 8,  
2 wherein said affinity surface is machined.

1 21. The nucleic acid extraction device of claim 8,  
2 wherein said affinity surface is injection molded.

1 22. The nucleic acid extraction device of claim 1,  
2 further comprising:  
3 a piezoelectric crystal adapted to acoustically agitate  
4 said sample.

1 23. A method for extracting nucleic acid from a sample  
2 comprising:  
3 positioning the sample in a miniature chamber having a  
4 structure with nucleic-acid binding properties disposed  
5 therein;  
6 binding nucleic acid from the sample to the structure;  
7 and  
8 drawing the sample from the miniature chamber.

1 24. The method for extracting nucleic acid from a  
2 sample as set forth in claim 22, wherein  
3 said structure is a porous fluid plug, and  
4 said binding step is accomplished by passing the sample  
5 through the structure.

1 25. The method for extracting nucleic acid from a  
2 sample as set forth in claim 22, further comprising the step  
3 of:

4 pretreating the structure with an agent for enhancing  
5 the nucleic acid binding properties.

1 26. The method for extracting nucleic acid from a  
2 sample as set forth in claim 22, wherein  
3 said agent is selected from the group consisting of  
4 acids, bases, silanes, polylysine, tethered antibodies, and  
5 Poly-T DNA.

1 27. A biological sample refinement device, comprising:  
2 a body having at least one microchamber with at least  
3 one inlet channel;

4 a structure disposed within the microchamber, the  
5 structure having binding sites thereon; and

6 a fluid distribution system for delivering a biological  
7 sample into the microchamber such that at least a portion of  
8 the sample contacts the binding sites.

1 28. The device of claim 27 wherein the binding sites  
2 are antibodies that are adhesively attached to the structure.

1 29. The device of claim 27 wherein the binding sites  
2 are oligonucleotides attached to the structure.

1 30. The device of claim 27 wherein the structure  
2 comprises a substantially planar wall with a plurality of  
3 beads attached thereto.

1 31. A deformable microchamber device, comprising:  
2 a pneumatic portion having an addressable port formed  
3 therein,  
4 a fluid portion having a reaction chamber formed  
5 therein,

6 said pneumatic portion and said fluid portion being  
7 bonded together with said addressable port being positioned in  
8 mating contact over said reaction chamber, and  
9 a deformable member disposed between said pneumatic  
10 portion and said fluid portion, said deformable member acting  
11 as a flexible chamber wall which seals the reaction chamber.

12 32. A method of forming a molded microcapillary,  
13 comprising the sequential steps of:  
14 forming a mold part,  
15 depositing a first parylene layer on a substrate part,  
16 affixing said mold part to said substrate,  
17 depositing a second parylene layer on said mold part  
18 and said substrate,  
19 removing said mold part from said substrate.

1 33. The method of forming a molded microcapillary in  
2 claim 32, wherein:  
3 said step of depositing a second parylene layer is  
4 accomplished by depositing parylene into cavities on said mold  
5 part.

1 34. The method of forming a molded microcapillary in  
2 claim 32, wherein:  
3 said step of removing said mold part from said  
4 substrate is accomplished by dissolving a release layer coated  
5 on said mold part.

1 35. A hermetically sealed microfluidic system,  
2 comprising:  
3 a body having at least two reaction chambers connected  
4 by a fluidic channel disposed therebetween,  
5 a pneumatic port connected to said chamber, said  
6 pneumatic port having a gas-liquid separator disposed therein,  
7 a pneumatic line, and  
8 a deformable diaphragm sealing said pneumatic port from  
9 said pneumatic line.

1 36. The hermetically sealed microfluidic system as set  
2 forth in claim 35, wherein:

3 said gas-liquid separator is a porous hydrophobic vent.

1 37. The hermetically sealed microfluidic system as set  
2 forth in claim 35, wherein:

3 said deformable diaphragm is selected from the group  
4 consisting of latex, polyimide, polypropylene, and mylar.

1 38. The hermetically sealed microfluidic system as set  
2 forth in claim 35, wherein:

3 said deformable membrane covers said gas-liquid  
4 separator.

1 39. The hermetically sealed microfluidic system as set  
2 forth in claim 35, further comprising:

3 a pneumatic manifold connected to said second pneumatic  
4 port at each of said at least one reaction chambers.

1 40. The hermetically sealed microfluidic system as set  
2 forth in claim 35, further comprising:

3 a pneumatic driving chamber connected to said pneumatic  
4 port, said pneumatic driving chamber having a displaceable  
5 pneumatic driving chamber vent for inducing pressure changes  
6 in said pneumatic port.

1 41. A microfluidic particle suspension valving  
2 arrangement, comprising:

3 a flow chamber having a narrow hydrophobic region,  
4 a particle emulsion disposed in said narrow region,  
5 said particle emulsion being immiscible in water, and  
6 generally occluding said narrow hydrophobic region.

1 42. The microfluidic particle suspension valving  
2 arrangement of claim 41, wherein

3 the viscosity of said particle emulsion can be varied  
4 by a magnetic field.

1 43. The microfluidic article suspension valving  
2 arrangement of claim 41, wherein  
3 the viscosity of said particle emulsion can be varied  
4 by an electric field.

1 44. In a ~~microfluidic~~ fluid system, an enzymatic  
2 reaction selected ~~selected~~ from the group consisting of terminal deoxy-  
3 transferase, DNAase, in vitro translation, and ligation.

1 45. A low-volume hybridization chamber, comprising:  
2 a base,  
3 a reaction chamber disposed in said base, said reaction  
4 chamber being bound by a flexible diaphragm, and  
5 a probe array disposed in said reaction chamber.

1 46. The low-volume hybridization chamber of claim 45,  
2 wherein  
3 said reaction chamber has a volume in the range of 0.1  
4 to 100 $\mu$ l.

1 47. The low-volume hybridization chamber of claim 45,  
2 wherein  
3 said reaction chamber has a volume in the range of 1 to  
4 20 $\mu$ l.

1 48. The low-volume hybridization chamber of claim 1,  
2 further comprising:  
3 a pneumatic system for moving said flexible diaphragm.

4 49. A hybridization device, comprising:  
5 a base,  
6 a fluidic chamber disposed in said base, said fluidic  
7 chamber having a hybridization array disposed therein,  
8 a porous membrane disposed in said fluidic chamber  
9 opposite said array,  
10 a pneumatic port disposed in said base, said pneumatic  
11 port addressing said porous membrane, and

12 a thermal control device for controlling the  
13 temperature in the array.

1 50. A miniature genetic analysis system comprising:  
2 a body having at least one reaction chamber disposed therein;  
3 an addressable heater adjacent to or within each  
4 chamber;  
5 a thermal insulation in contact with said heater;  
6 a cooler coupled to said thermal insulator and disposed  
7 to cool each of the reaction chambers;  
8 a temperature sensor positioned adjacent said heater;  
9 and  
10 a temperature controller.

1 51. The system of claim 50 wherein the insulator  
2 comprises a polymeric film having a thickness of about 0.1 mm  
3 to about 1.0 mm.

1 52. A method for linking together two spaced-apart  
2 fluid plugs disposed in a first capillary tube, wherein said  
3 first capillary tube intersects a second capillary tube having  
4 a gas-liquid separator extending therefrom, comprising:  
5 moving said first fluid plug along said first capillary  
6 tube such that a leading edge of said first fluid plug moves  
7 into said second capillary tube and reaches said gas-liquid  
8 separator with a trailing edge of said first fluid plug  
9 remaining in said first capillary tube,  
10 forcing gas through said gas-liquid separator thereby  
11 expelling fluid from said second capillary tube, and  
12 moving a second fluid plug along said first capillary  
13 tube towards said leading edge of said first fluid plug tube  
14 such that a leading edge of said second fluid plug moves into  
15 said second capillary tube with a trailing edge of said second  
16 fluid plug remaining in said first capillary tube.

1 53. A device for removing gas bubbles and linking  
2 together fluid plugs in a microfluidic system, comprising:



3 an elongated chamber having a wide portion and a narrow  
4 portion,  
5 a first input port opening into the narrow portion of  
6 said elongated chamber, and  
7 a gas exhaust port opening into the wide portion of  
8 said elongated chamber.

1 54. The device for removing gas bubbles and linking  
2 together fluid plugs in a microfluidic system as set out in  
3 claim 53, further comprising:  
4 a second input port opening into the wide end of said  
5 elongated chamber.

1 55. The device for removing gas bubbles and linking  
2 together fluid plugs in a microfluidic system as set out in  
3 claim 53, wherein:  
4 said elongated chamber has a narrowed width portion  
5 extending along its longitudinal length.

1 56. A method for removing gas bubbles and linking  
2 together fluid plugs in a microfluidic system, comprising:  
3 exerting a pressure differential to move a capillary  
4 stream consisting of spaced apart fluid plugs with gas bubbles  
5 inter-disposed therebetween into a narrow portion of an  
6 elongated chamber, and  
7 removing said gas bubbles from said elongated chamber  
8 through a port connected to a wide portion of said elongated  
9 chamber, wherein said wide portion is positioned opposite  
10 said narrow portion.

1 57. A method for removing gas bubbles and linking  
2 together fluid plugs in a microfluidic system, comprising:  
3 exerting a pressure differential to move a capillary  
4 stream consisting of spaced apart fluid plugs with gas bubbles  
5 inter-disposed therebetween into a wide end of an elongated  
6 chamber, and

7 removing said gas bubbles from said elongated chamber  
8 through a port connected to a narrow end of said elongated  
9 chamber, wherein said wide end is positioned opposite said  
10 narrow end.

1 58. A device for manipulating nucleic acids in a  
2 sample, comprising:  
3 a base defining a reaction chamber,  
4 a first chamber extending from said reaction chamber,  
5 said first chamber having a first electrode received therein,  
6 a second chamber extending from said reaction chamber,  
7 said second chamber having a second electrode received  
8 therein, and  
9 a first barrier disposed between said reaction chamber  
10 and said first chamber, and  
11 a second barrier disposed between said extraction  
12 chamber and said second chamber.

1 59. A microfluidic controlled pH device, comprising:  
2 a reaction chamber,  
3 a first and second electrode disposed in said reaction  
4 chamber,  
5 a counter-electrode chamber in fluid connection with  
6 said reaction chamber, said counter-electrode chamber and said  
7 reaction chamber having a barrier disposed therebetween, and  
8 a fourth electrode.

1 60. A microfluidic acoustic treatment device,  
2 comprising:  
3 a chamber having formed in a polymeric base, said  
4 chamber having a lower surface with a plurality of  
5 microstructures formed therein and a thin upper wall,  
6 an acoustic source coupled to said reaction chamber.

1 61. A device for acoustic manipulation of biological  
2 particles, comprising:

an array of transducers for producing acoustic standing waves.

62. The device for acoustic manipulation of biological particles of claim 61, wherein:

said transducers comprise surface-acoustic wave transducers.

63. The device for acoustic manipulation of biological particles of claim 61, wherein:

said transducers comprise flexural plate wave transducers.

64. A method of providing a measured dose of fluid into a common line in a capillary system, comprising:

pressurizing a common line to cause a fluid plug to enter a sealable chamber intersecting said common line, holding the fluid plug in said sealable chamber by closing a valve positioned on said sealable chamber proximal the intersection of said sealable chamber and said common line,

evacuating said common line, and opening said valve to permit a measured dose of fluid to move from said sealable chamber to said common line.

65. A device for linking fluid plugs in a microfluidic system, comprising:

a first capillary tube having two valves positioned therealong, and

a second capillary tube extending from said first capillary tube and having a gas-liquid separator positioned therealong.